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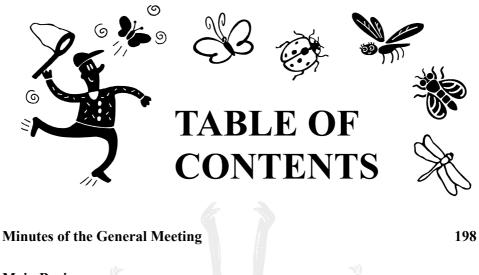
The ENTOMOLOGICAL SOCIETY OF QUEENSLAND INC., since its inception in 1923, has striven to promote the development of pure and applied entomological research in Australia, particularly in Queensland. Membership is open to anyone interested in Entomology. The Society promotes liaison among entomologists through regular meetings and the distribution of a *News Bulletin* to members. Meetings are announced in the *News Bulletin*, and are normally held on the second Monday of each month (March to June, August to December), or on Tuesday if Monday is a public holiday. Visitors and members are welcome. Membership information can be obtained from the Honorary Secretary, or other office bearers of the Society.

Contributions to the *News Bulletin* such as items of news, trip reports, announcements, etc are welcome and should be sent to the News Bulletin Editor.

The Society publishes **THE AUSTRALIAN ENTOMOLOGIST**. This is a refereed, illustrated journal devoted to Entomology in the Australian region, including New Zealand, Papua New Guinea and the islands of the South Western Pacific. The journal is published in four parts annually.

EMBLEM: The Society's emblem, chosen in 1973 on the 50th anniversary of the Society, is the king stag beetle, *Phalacrognathus muelleri* (Macleay), family Lucanidae. Its magnificent purple and green colouration makes it one of the most attractive of all Australia Coleoptera. It is restricted to the rainforests of northern Queensland.

COVER: *Diadegma semiclausum* ovipositing into a larva of the diamondback moth, *Plutella xylostella*. Drawn by Sandra Dennien.



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The issue of this document does **NOT** constitute a formal publication for the purposes of the "International Code of Zoological Nomenclature 4th edition, 1999". Authors alone are responsible for the views expressed.

<u>The Entomological Society of Queensland</u> <u>Minutes of General Meeting 10 November, 2008</u>

Held in the Large Conference Room, CSIRO Entomology, Long Pocket Labs, 120 Meiers Road, Indooroopilly, on 10 November, 2008, 12.00 midday.

The meeting was chaired by Senior Vice President, Christine Lambkin.

Attendance:

Richard Bull, Chris Lambkin, Noel Starick, Geoff Monteith, Geoff Thompson, Don Sands, Susan Wright, Anna Marcora, Shaun Winterton, Tim Heard, Desley Tree, Ross Kendall, Lynita Howie, Belinda Walters, Felix Bianchi, Peter Gillespie.

Visitors:

Lloyd Dosdall, Karen Bell, Anne Bourne, Nicole Reid, Brendan Murphy, Juan A.Villanueva-Jimenez, Holger Loecker, Nate Hardy.

Apologies:

Peter Allsopp, Matthew Purcell, Lyn Cook, Gunter Maywald, Geoffrey Shaw, John Moss, John Goolsby, Bill Palmer, Hilton Selvey, Stacey McLean, Penny Mills, Graham Forbes.

Minutes:

The minutes of the last General Meeting of October 13th, were circulated in the News Bulletin Vol. 36, Issue 7, 2008.

Moved the minutes be approved: Don Sands *Seconded:* Geoff Thompson

Membership Nominations and Elections:

Dr Dennis Bell. The nomination was announced and approved by a unanimous show of hands.

General Business:

Don Sands addressed the audience and advised he was required to re-write the format of Collecting Permits for Protected Species and after lengthy consultation with EPA, would have it ready for early 2009. He advised there would be new, restrictive conditions for its use. Main Business:

<u>The landscape context of the ecosystem</u> <u>service of pest control</u>

Dr Felix Bianchi - CSIRO Entomology

Natural pest regulation is an important ecosystem service with an estimated value of several hundreds of billion dollars per year at a world-wide scale. Due to the activity of natural enemies the vast majority of potential arthropod pest species are controlled and do not reach outbreak levels in forest and agro-ecosystems. A sound natural pest regulation function can therefore contribute to a reduction of pesticide use and the associated adverse environmental effects.

There is increasing evidence that the effectiveness of natural enemies to suppress pest populations in crops depends on landscape context. In Australia, approximately 6% of the land area is used for cropping in regions with favourable rainfall. Although is only a small fraction of the continent, there have been dramatic changes in these landscapes, including clearing of native vegetation and expansion of the area in which crops are grown year round. As a result, intensively used agricultural areas are often merely composed of large-scale monocultures of only few crops types, with only little or no native vegetation left. These changes in land use may have affected the population dynamics and interactions of pest insects and natural enemies.

Habitats in agricultural landscapes can have various functions for pest and natural enemies. Crops often provide excellent host plants for herbivorous insects because of fertilization and irrigation. However, crops are also ephemeral and highly disturbed habitats, for instance by pesticide application and harvesting. The intense and frequent disturbances make crops hostile environments for many natural enemy species. In contrast, native vegetation usually persist at the same place for a long time and may therefore continuously provide resources for herbivorous insects and natural enemies. Native vegetation may be crucial habitats for many natural enemies because they provide alternative prey or hosts when pest densities in crops are low. Native vegetation may also provide floral food resources, such as nectar and pollen, which are essential for adult stages of parasitoids, hoverflies and lacewings. Presence of floral food sources can enhance the longevity and reproductive output of parasitoids, thus increasing their effectiveness as biological control agents. Native vegetation can also provide a moderated micro-climate as opposed to crops. Laboratory studies have demonstrated that the longevity of parasitoids at high temperatures is clearly lower than at moderate temperatures. Native vegetation can therefore potentially act as refuge areas during the heat of the day. Finally, native vegetation is also associated with non-insect species that can provide pest control services, such as insectivorous bats.

Despite the clear indications that presence of native vegetation in agricultural landscapes may influence natural enemy populations and the ecosystem service of pest control, there have been only a very limited number of Australian studies focussing on this issue. Studies have shown that natural enemies are present in native vegetation and that they move between native vegetation and crops. However, it is not clear to what extend native vegetation acts as a reproduction (source) habitat which can generate natural enemies that can provide pest control services in the surrounding landscape. Nor do we know how far these pest control services are extending from source habitats. In a study funded by the Cotton Catchment Communities CRC and Land and Water Australia the role of native vegetation for sustaining the ecosystem service of pest control is investigated. The aim of the study is twofold: (i) the identification of source habitats for pests and natural enemies, and (ii) the spatial scale at which native remnants contribute to the ecosystem of pest control.

The study is conducted in two 10 km diameter landscapes in the Darling Downs region near Dalby. One landscape is characterized by arable fields intermingled with patches of native remnants and tree lines, whereas the other landscape contained only a few tree lines (Fig. 1 and 2).

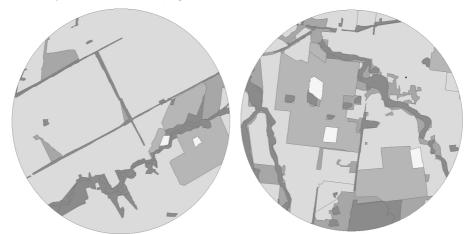


Fig. 1. Map of the two experimental landscapes. Water is indicated in white, cropping (irrigated and dry land) in light grey shades, and native vegetation in dark grey.

For our first objective, i.e. the identification of source habitats for pests and natural enemies, crops and native vegetation was sampled in October 2007, January 2008 and early April 2008 to assess pest and natural enemy densities. We were particularly interested in the presence of immature stages because this indicates that habitats may function as a source for pests or natural enemies. Crops were sampled by beat sheet (6 repetitions per field) and a plant sample was taken and put in polypropylene bags to rear out pests and/or natural enemies. The assessment of native vegetation focussed on poplar box (Eucalyptus populnea), an acacia species (Acacia salicina) and several sultbushes (Enchylaena tomentosa, Atriplex muelleri, Sclerolaena muricata, Rhagodia nutans and Maireana microphylla). These species were sampled by beat sheet (3 repetitions per species) and DVAC, and plant clippings were taken to rear out pests and/or natural enemies. Our findings showed that native vegetation can act as a strong source habitat of natural enemies during spring and typically do not produce high pest numbers (Fig. 3). In contrast, many crop types act as source habitats of pests, and much less so for natural enemies. This suggests that native vegetation has a positive effect on the natural enemy : pest ratio at the landscape scale, and that native vegetation may act as a green bridge between spring and summer crops or drought conditions aiding colonisation of natural enemies into crops.



Fig. 2. Native vegetation next to an arable field.

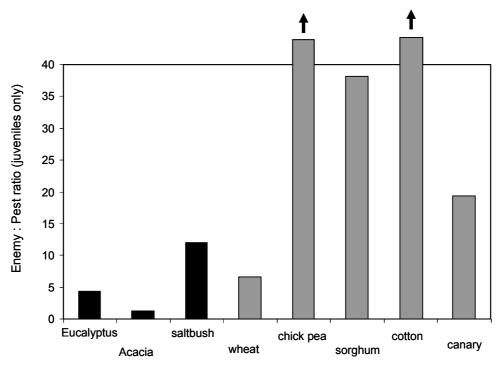


Fig. 3. Natural enemy : pest ratio of juvenile stages in native vegetation (black) and crops (grey).

For our second objective, i.e. assessment of the spatial scale at which native remnants contribute to the ecosystem of pest control, modules with experimental cotton seedlings were set out in native vegetation, bare arable fields adjacent to native vegetation, and bare arable fields that were at least 400 m from native vegetation. In total six native vegetation remnants and twelve fields were studied. Fields adjacent and far from native vegetation received 20 modules, whereas remnants received 12 modules (Fig. 4). Modules were placed in grids with a 4 x 5 design (fields adjacent and far from native vegetation) or a 4 x 3 grid design (native vegetation). The modules consisted of cotton seedlings containing (i) cards with Helicoverpa eggs, (ii) whitefly nymphs, and (iii) no insects (i.e. clean plants; Fig 5 and 6). The pest infested seedlings were used to assess the rate at which natural enemies remove/parasitize pests, whereas the clean seedlings were used to quantify the rate of pest colonization. In addition, sticky traps were used to sample the insect community around the seedlings. The cotton seedlings stayed out for three full days before they were collected from the fields and remnants. The experiment was repeated three times (13-15 October, 17-20 October and 21-24 October).

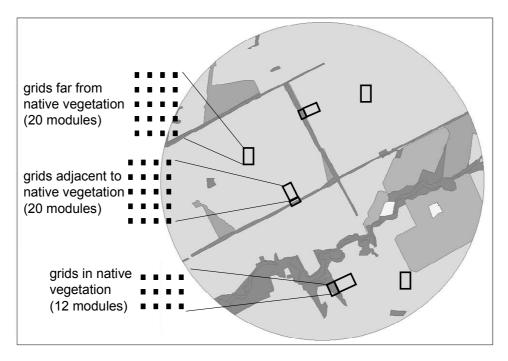


Fig. 4. Map with grids far, adjacent and in native vegetation indicated.



Fig. 5. Setting out of the modules.



Fig. 6. Module containing cotton seedlings with (i) whitefly nymphs, (ii) cards with *Helicoverpa* eggs, (iii) jassid eggs (data not presented), (iv) no insects (i.e. clean plants), as well as a sticky trap.

Parasitism of cotton plants with whitefly nymphs was significantly higher in native vegetation and fields adjacent to native vegetation than fields far from native vegetation, indicating that native vegetation can act as a source of whitefly parasitoids (Fig. 7). In contrast, predation rates of Helicoverpa eggs on cotton plants were not significantly different among the treatments (Fig. 8). We observed that ants were very active in removing eggs from the plants in fields far and close to native vegetation, as well as in native vegetation. As for pest colonisation, no effects of native vegetation were found for jassids (Fig. 9A), suggesting that native remnants do not play a particularly important role for jassids. Aphids were hardly found outside native vegetation (Fig. 9B). This suggests that although aphid densities in native remnants may be high, they do not pose a threat to cotton crops early in the growing season. Whiteflies typically colonized sentinel plants in the fields (Fig. 9C), but not in native vegetation, showing that native vegetation is not a source for whiteflies. For Helicoverpa spp., there was a trend of higher colonisation of sentinel plants in native vegetation (Fig. 9D), but this was not statistically significant. Interestingly, we hardly found Helicoverpa spp. during our sampling on poplar box, acacia and saltbush, suggesting that *Helicoverpa* adults are ovipositing on cotton when introduced in native vegetation, but not on native vegetation itself.

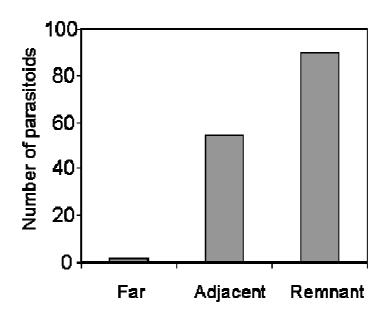


Fig. 7. Number of whitefly parasitoids in native vegetation (remnant), fields adjacent to native vegetation and fields far from native vegetation.

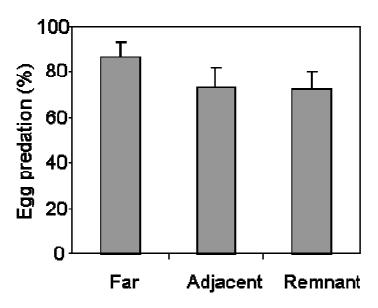
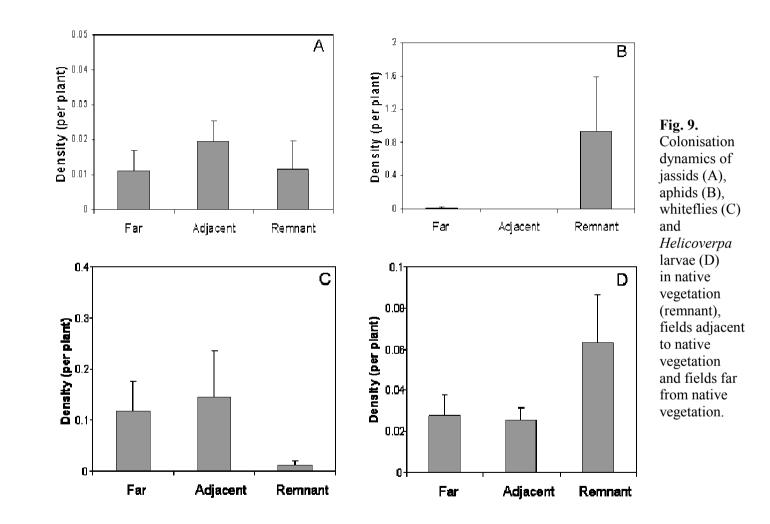


Fig. 8. Egg predation rates in native vegetation (remnant), fields adjacent to native vegetation and fields far from native vegetation.



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In conclusion, our data demonstrate that native remnants can play a crucial role in capturing the ecosystem service benefit of pest control. On-farm conservation of biodiversity by managing existing native vegetation or revegetation can therefore contribute to more sustainable cotton production systems, which are less reliant on the use of chemical pesticides. Future research will help us to further understand how pests and natural enemies are using crops and native vegetation, and will help us to design cropping systems for multiple benefits including better pest and pollination services, enhanced biodiversity, and profitability.



Vote of thanks was given by Richard Bull.

Chairman's closing statement:

The next meeting will be held at this venue on Monday 8 December at 5.00pm with Notes and Exhibits followed by a "Sausage Sizzle" at 6.00pm (\$5 per person). Drinks will also be available for purchase. Please RSVP for catering purposes and also advise of any special dietary requirements - Contact Matt Purcell on 3214 2847.

Notes & Exhibits:

Geoff Thompson – A piece of entomological sporting memorabilia.

Chris Lambkin & Noel Starick – 'Raining' trees.

Desley Tree – Diet & feeding method of *Mecynothrips hardyi*, a spore feeding thrips.

Additional Notes & Exhibits are welcome. These should be brief and informal items of interest. Please contact the President if you wish to make a presentation (contact details can be found on the back page of the News Bulletin).

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Notice of Next Meeting				
<u>Entomological Society</u> <u>of Queensland</u>				
Monday 8th December 2008 <u>5pm</u>				
CSIRO Long Pocket Laboratories Large Conference Room 120 Meiers Rd, Indooroopilly Notes & Exhibits				
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A piece of entomological sporting memorabilia.				
<u>Christine Lambkin & Noel Starick</u> 'Raining' trees. <u>Desley Tree</u> Diet & feeding method of <i>Macynothrins hardvi</i>				
<u>Christine Lambkin & Noel Starick</u> 'Raining' trees. <u>Desley Tree</u> Diet & feeding method of <i>Mecynothrips hardyi</i> , a spore feeding thrips.				
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Notes and Exhibits will be followed by a <u>"Sausage Sizzle" at 6.00pm</u> Cost: \$5 per person and drinks will also be available for purchase. Please RSVP for catering purposes and also advise of any special dietary requirements - Contact Matt Purcell on 3214 2847.				
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<u>News from the USDA-ARS Australian Biological</u> <u>Control Laboratory</u>

Jeff Makinson and Tony Wright visited Hong Kong with our USDA ARS collaborator, Dr. Ted Center. They conducted surveys for biological control agents of *Rhodomyrtus tomentosa* and *Lygodium microphyllum* in the New Territories. Tony then accompanied Dr. Center to Thailand where they undertook further surveys. Jeff Makinson joined Matthew Purcell in Wuhan, China where they met with staff from the Chinese Academy of Sciences and Dr. Greg Wheeler (USDA ARS). Joint surveys were conducted throughout southern China looking for biological control agents for *Hydrilla verticillata*, *Lygodium microphyllum* and *Triadica sebiferum*.

News Bulletin contributions from Entsoc Members We would love to receive your news, field trip reports, sightings of strange and wonderful beasts, Entomological Notes, Bug of the Month, gossip, concerns, questions and suggestions pertaining to the world of entomology. Please send contributions to the News Bulletin editor or your nearest Entsoc office bearer. Don't delay, next issue out soon! Thank you, Anna

Record-breaking insect goes to extraordinary lengths

The world's longest insect has been revealed by the Natural History Museum. Measuring more than half a metre, the length of this stick-insect is almost as long as your arm.

The thin-bodied creature, which goes on display today in the Museum Creepy Crawlies gallery, has just been named *Phobaeticus chani* (Chan's megastick) and is an incredible 56.6 centimetres with its legs fully stretched. This is more than one centimetre longer than the former record holder for overall length, a stick-insect called *Phobaeticus serratipes* found in Malaysia and Indonesia. Without including its legs, the new species measures 35.7 centimetres, winning the insect world record for the longest body and beating the previous title-holder, *Phobaeticus kirbyi* from Borneo, by 2.9 centimetres.

'We've known about both the previous record holders for over 100 years, so it's extraordinary an even bigger species has only just been discovered,' said Dr George Beccaloni, curator of stick-insects and their relatives at the Natural History Museum. 'It is a sad thought that many other spectacular insect species are disappearing as their habitats are destroyed, before we have had the chance to find and name them.'

Although virtually nothing is known about the biology and ecology of this supersized insect, it is thought it probably lives in the canopy of the rainforest, making it especially hard to find. In addition to its size, its eggs may also be unique in the insect world. Each egg capsule has wing-like extensions on either side like a miniature golden snitch (the flying sports ball in the *Harry Potter* books), allowing them to drift in the wind when the female drops them, therefore helping the species spread.

There are around 3,000 known species of stick-insect, mainly living in the tropics and subtropics. Although they do not occur naturally in Britain, three species from New Zealand have become established in the southwest of England and the Isles of Scilly.

Only three specimens of the new insect have so far been found, all from the Malaysian state of Sabah on the island of Borneo. The insect's namesake, Datuk Chan Chew Lun from Sabah, obtained the first and largest known specimen from a local collector and has donated it to the Natural History Museum. The other two specimens are in collections in Sabah. British scientist Dr Philip Bragg described and named this stick-insect for the first time this week and details are in the journal *Zootaxa*. The Entomology Department is one of six science departments at the Natural History Museum. The Entomology collections amount to 28 million specimens stored in 140,000 drawers. Scientists in the department study insects and other terrestrial arthropods, including spiders and mites in a wide range of research projects across the world.



16 October 2008

http://www.nhm.ac.uk/about-us/news/2008/october/worlds-longest-insect-revealed.html



Giant stick insect - *Phobaeticus chani* - and curator George Beccaloni (Copyright NHM)



Phobaeticus chani (Copyright NHM)

<u>Numbers of Living Species in Australia</u> <u>and the World</u>

Call for information for the 2nd edition

ABRS is requesting input for the 2nd edition of the Numbers of Living Species in Australia and the World.

In 2006 ABRS published a report titled *Numbers of Living Species in Australia and the World*. The report was compiled by Arthur Chapman, and was made available in hardcopy and online. It provided a summary and further discussion on numbers of species by group.

The online version of the publication (html & pdf) is available at:

http://www.environment.gov.au/biodiversity/abrs/publications/other/ species-numbers/index.html

This document was largely a synthesis of published information, and information supplied to Arthur by researchers around Australia and the world, and Arthur would like to thank all those who supplied him with information at the time.

ABRS is now preparing to have the document updated and if anyone has any comments on the document, references to relevant papers, or any other information, we would be most grateful if you could send this on to Arthur for use in the 2nd edition.

Following feedback from the 1st edition Arthur would like to split up some groups a bit more if possible - for example the insects to order level if information on the numbers in each order is available (please see the document to see the current groupings at):

http://www.environment.gov.au/biodiversity/abrs/publications/other/ species-numbers/index.html

All comments and references will be fully acknowledged as in the 1st edition.

Please send any information to Arthur Chapman at the address below: Arthur D. Chapman (Trading as: Australian Biodiversity Information Services) PO Box 7491, Toowoomba South, Qld 4352 Australia Home Phone: International: +61 7 4630 9065 Mobile: International: +61 400 400 326 Skype: Kowari47 Email: biodiv_2@achapman.org

Flies land on Mackerras Medal

CSIRO Media Release: 28 October 2008 http://www.csiro.au/news/MackerraMedal08.html

CSIRO Entomology scientist, Dr David Yeates, has been awarded the Australian Entomological Society's 2008 Mackerras Medal.

"Dr Yeates' award recognises his enormous input into understanding the taxonomy, systematics and evolution of flies as well as his extensive support for Australian taxonomy in general," said Dr Mark Lonsdale, Chief CSIRO Entomology.

"Since becoming curator of Diptera (flies) at the Australian National Insect Collection (ANIC) in 2001, Dr Yeates has reinvigorated fly research."

"Now his research position in ANIC is funded in perpetuity by an endowment from the United States based Schlinger Foundation."

Dr Lonsdale said Dr Yeates enjoys a very high standing in the international dipterological community and is a collaborator, advisor or reviewer in almost all major international collaborations in his areas of expertise.

These include a 10-year project on the world's stiletto flies (Therevidae) and a five-year project on the Diptera Tree of Life. This project is part of the much larger US National Science Foundation's *Assembling the Tree of Life*.

Dr Yeates is currently President of the Council of the International Congresses of Dipterology, Vice President of the Australian Entomological Society and a Co-Principal Investigator on the Australian Government's Commonwealth Environment Research Facilities (CERF) funded Taxonomy Research and Information Network (TRIN).

As well as an impressive publication history on almost all aspects of fly taxonomy, systematics and evolution, Dr Yeates was part of the team which won The Royal Zoological Society of NSW 2007 Whitley Book Awards Certificate of Commendation for the interactive CD, *On-The-Fly*.

Before joining CSIRO, he was a highly successful academic at the University of Queensland.

The Mackerras Medal is the Australian Entomological Society's highest award and is given every two years to a member of the Society under 50 years of age who has demonstrated excellence in entomology.

It is named in honour of Dr Ian Mackerras, the Society's first President and an entomologist who embodied the excellence in entomological pursuits that the award recognises.



Above: A fly from the Family Pelechorhynchidae - nectar and pollen feeders. Image Credit - David McClenaghan, CSIRO.



Right: A fruit fly. Image Credit -CSIRO.

Pollinator decline not reducing crop yields just yet

CSIRO Media Release: 10 November 2008 http://www.csiro.au/news/Pollinator-Decline.html

The well-documented worldwide decline in the number of bees and other pollinators is not, at this stage, limiting global crop yields, according to the results of an international study published in the latest edition of the respected science journal, Current Biology.

Co-author, CSIRO Entomology's Dr Saul Cunningham, says however that the study detected warning signs that demand for pollinators is still growing and some highly pollinator-dependent crops are suffering.

"The research team scored crops on how much they depend on pollinators for maximum production," Dr Cunningham says. "Depending on the crop, this dependence ranges from zero to 100 per cent. For example, cereal crops like wheat don't need to be pollinated but at the other end of the scale, unpollinated almond trees produce no nuts."

The team found that between 1961 and 2006 the yields of most crops have consistently grown at about 1.5 per cent a year because of improvements in agriculture. There was also no difference in yield growth between crops that require pollinators and those that do not.

"While this is a positive finding, the interaction between yields and pollination is a hugely complex issue which needs to be teased-out further," Dr Cunningham says. "Global summaries can also hide local stories. In some places, local pollinator shortages are affecting local production. While these don't threaten overall global food supplies, they can have very significant impacts on local communities and their economies."

The researchers were surprised to discover that there has been a global increase in the growing of pollinator-dependent crops, particularly in the developing world. "The fact that, while pollinators are declining in various parts of the world, global agricultural systems are becoming more dependent on pollinators, could create serious problems in the future," Dr Cunningham says.

When the group looked at pollinated crops in more detail, they found that pollinator shortages might be beginning to affect crops that rely heavily on pollination because their yield growth was lower than for crops that were less dependent.

Dr Cunningham says they now want to examine how declining pollinator supply might be increasing the costs of production. Increased yields are usually the result of increasing farm inputs such as fertiliser, labour and water. For some crops, this increasingly intensive management may have, for now, overcome any losses in pollinator service, but it also increases production costs. There is also evidence that one response to lower yield growth for highly pollinator dependent crops is a growing demand for land.



There is as yet no evidence that global pollinator decline is limiting global food production. Image credit - Sujaya Rao, CSIRO.

<u>Nominations for 2009 Office Bearers of the</u> <u>Entomological Society of Queensland</u>

Members are invited to use the following form to nominate office bearers for the Entomological Society of Queensland Inc. for 2009.

Nominations should be sent via email, fax or post and be referred to the:

Secretary, Entomological Society of Queensland

PO Box 537, Indooroopilly QLD 4068

Please return forms by the end of January 2009.

A list of nominations received will be circulated in Issue 10 of the News Bulletin, and an election held at the Annual General Meeting in March 2009. In the absence of a nomination for any particular office, the president may receive nominations at the Annual General Meeting.

Positions to be filled are as follows:

- Senior Vice President
- Honorary Secretary
- Honorary Treasurer
- News Bulletin Editor
- Councillors (3 positions)

The Entomological Society of Queensland functions effectively because members play an active part in the Society. All members are encouraged to nominate for positions on the Council of the Society. If you want to know more about any of the Council positions, please contact one of the existing Council members listed on the back cover of the News Bulletin.

Office Bearer Nomination Form 2009

I nominate (name)

.....

For the position of

- Senior Vice President
- Honorary Secretary
- Honorary Treasurer
- News Bulletin Editor
- Councillor

on the Council of the Entomological Society of Queensland Inc.

Nominated by

.....

Seconded by

.....

I accept the nomination

(nominee signature)

Entomological Society of Queensland 2009 \$250 Student Award

This is an award by the Society to encourage entomological research. Entries are judged by a panel of 3 entomologists appointed by the President of the Society. The winner will be announced at the May General Meeting and is then invited to present a summary of their research at the June Notes and Exhibits meeting of the Society.

Honours, Diploma and 4th year Degree students who received their qualification from any Queensland tertiary education institution in 2008 or 2009 may submit their entomology based thesis or report for consideration.

Entrants need not be Society members.

These reports can be directed to the Society's Senior Vice President at the address listed on the entry form. However, please note that a hard copy of your thesis/report does not need to be submitted, and the submission of a PDF version is encouraged. This should be emailed together with a signed copy of the completed entry form to Christine Lambkin at christine.lambkin@qm.qld.gov.au

Closing date for submissions is Friday 17th April 2009.

Student Award Sponsors:

Tropical Fruit Fly Research Group, Griffith University



Entomological Society of Queensland 2009 Student Award Entry Form

Name			
Title of thesis or report			
Degree			
Supervisor			
Date of Examiners report or grading			
Return address for thesis/report (if applicable)			
Signature	Date		
Send in thesis/report with a signed and completed entry form to: Christine Lambkin Senior Vice President of the Entomological Society of Queensland Queensland Museum PO Box 3300, South Brisbane, QLD 4101 Fax: 07 38461226			

To: Users of the Entomological Society of Queensland QPWS collecting permits

From: President, Entomological Society of Queensland

Re: Use and conditions of collecting permits

Please note that you are required to follow the conditions as stated on the permit. The permit and membership to the ESQ may be revoked if conditions are not followed. In particular note that:

- you are required to submit a return of operations within 30 days after the end of each 12 month period that the permits are in force;
- specimens may not be given, sold or traded (but a member who is endorsed on the permit may collect specimens on behalf of another member provided that they are also endorsed);
- wildlife habitat must not be damaged, environmental impact must be kept to a minimum;
- this permit does not give you the right to enter any land, you must get prior permission from the agency responsible for administration of the land;
- where possible, collecting activities should be effected away from public view;
- a copy of any resulting reports/activities derived from this research should be provided to the EPA office at which the permit was issued;
- you must carry a copy of the permit that is endorsed by the permit holder (S.Wright) and it must contain your name and residential address;
- you must carry a form of identification that displays a coloured photograph such as a Queensland drivers license.

Complete and return the attached application form to:

Susan Wright Entomology Queensland Museum PO Box 3300 South Brisbane Qld 4101 Ph 07 3840 7704 Fax 07 3846 1226 Email: Susan.Wright@qm.qld.gov.au

Signed applications sent by fax are preferred.

<u>Application Form for use of The Entomological</u> <u>Society of Queensland Collecting Permits</u>

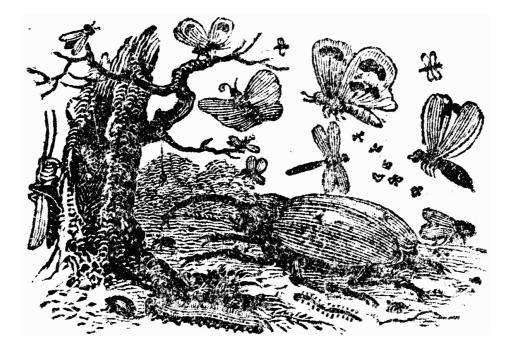
Name:
Residential Address:
Postal Address (if different from above):
Phone number:
Fax number:
Email address:
Project outline:
Parks you wish to visit:
Which permit will you be requiring? (please tick)
National Parks: State Forests:
Declaration: I agree that in using the permit I will read all conditions and procedures relating to these permits and will abide by them.
Signed :

<u>Note to users of the Entomological Society of</u> <u>Queensland insect collecting permits</u>

Information on the new insect collecting permits will soon be available on the website with a lot of tips and extra information for members who are not familiar with the new system.

Society's Web Address:

www.esq.org.au



DIARY DATES 2008					
Meetings held 2nd Monday of the month (or Tuesday if Monday is a public holiday)					
August 11th	Dr Peter James (Qld DPI&F)	Lousy research & the Integrated Parasite Management Group			
September 8th	Dr Shaun Winterton (Qld DPI&F)	Evolution of the Mantid lacewings based on multiple genetic markers (Neuroptera: Mantispidae)			
October 13th	Mr Stacey McLean	Filling Biodiversity Information Gaps in Brisbane City: Bugs, Birds and Bracket Fungi			
November 10th	Dr Felix Bianchi (CSIRO Entomology)	The landscape context of the ecosystem service of pest control			
December 8th	Notes & Exhibits				

IMPORTANT NOTICE

The official address for the Entomological Society of Queensland and *Australian Entomologist* and to which all communications should be addressed is: **PO Box 537, Indooroopilly QLD 4068.**

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NOTICE OF NEXT MEETING

The next meeting of the Society will be held at 5:00 pm on Monday, 8th December 2008 in the Large Conference Room, CSIRO Long Pocket Laboratories, 120 Meiers Rd Indooroopilly. The main business will be Notes & Exhibits: Geoff Thompson (A piece of entomological sporting memorabilia), Christine Lambkin & Noel Starick ('Raining' trees) & Desley Tree (Diet & feeding method of Mecynothrips hardyi, a spore feeding thrips). Additional Notes & Exhibits are welcome. Notes and Exhibits will be followed by a "Sausage Sizzle" at 6.00pm (\$5 per person).

VISITORS ARE WELCOME (Please sign in at CSIRO Reception before attending the meeting)

HONORARY LIFE MEMBERS OF THE SOCIETY

R.A.I. Drew D.L. Hancock M.J. Harslett D.S. Kettle D.P.A. Sands R.P. Kleinschmidt

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